

## Novel Human Histamine H<sub>3</sub> Receptor Antagonists

Chandra Shah, Laura McAtee, J. Guy Breitenbucher, Dale Rudolph, Xiaobing Li, Timothy W. Lovenberg, Curt Mazur, Sandy J. Wilson and Nicholas I. Carruthers\*

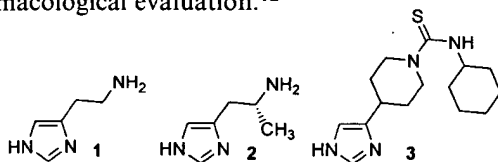
Johnson and Johnson Pharmaceutical Research and Development, L.L.C., 3210 Merryfield Row, San Diego, CA 92121, USA

Received 1 July 2002; accepted 15 August 2002

**Abstract**—High throughput screening, using the recombinant human H<sub>3</sub> receptor, was used to identify novel histamine H<sub>3</sub> receptor antagonists. Evaluation of the lead compounds ultimately afforded potent, selective, orally bioavailable compounds (e.g., **38**) with favorable blood–brain barrier penetration.

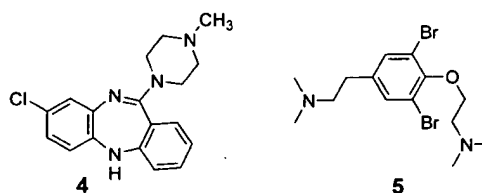
© 2002 Elsevier Science Ltd. All rights reserved.

The monoamine histamine (**1**) exerts a physiological effect via four distinct G-protein coupled receptors. Thus histamine plays a role, via the H<sub>1</sub> receptor, in immediate hypersensitivity reactions upon IgE mediated release from mast cells.<sup>1,2</sup> Histamine is also an important regulator of gastric acid secretion through its action upon H<sub>2</sub> receptors expressed in parietal cells.<sup>3</sup> The third histamine receptor (H<sub>3</sub>) identified in 1983, was first described as a pre- and postsynaptic autoreceptor in the brain<sup>4</sup> and subsequently as a presynaptic heteroreceptor on non-histamine containing neurons in both the central and peripheral nervous system.<sup>5</sup> Recently, a fourth histamine receptor (H<sub>4</sub>) was described with an expression profile that suggests a role in immune function.<sup>6–10</sup> Our interests relate to ligands for the histamine H<sub>3</sub> receptor, in particular histamine H<sub>3</sub> receptor antagonists, and follows from the successful cloning of the human H<sub>3</sub> receptor.<sup>11</sup> Histamine H<sub>3</sub> ligands are postulated to have a range of therapeutic applications and both H<sub>3</sub> agonists (e.g., *R*- $\alpha$ -methylhistamine **2**) and antagonists (e.g., thioperamide **3**) have undergone pharmacological evaluation.<sup>12</sup>

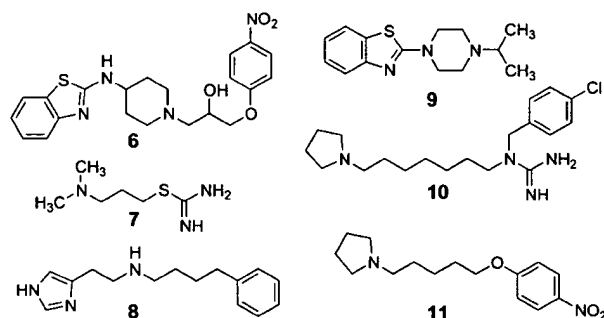


Until recently, a common feature of these molecules is their retention of the imidazole nucleus present in the

natural ligand. This results in compounds with poor blood–brain barrier (BBB) penetration<sup>13</sup> and metabolic liabilities associated with the imidazole ring.<sup>14</sup> Although compounds such as clozapine<sup>15</sup> **4** and the marine natural product aplysinine<sup>16</sup> **5** were reported to be H<sub>3</sub> antagonists, earlier efforts to identify imidazole replacements were disappointing.<sup>17</sup>



Only very recently have several groups described series of potent non-imidazole based histamine H<sub>3</sub> antagonists. Thus these groups have used low affinity H<sub>3</sub> ligands sabeluzole<sup>18</sup> **6**, dimaprit<sup>19</sup> **7** and *N*<sup>α</sup>-(4-phenylbutyl)histamine<sup>20</sup> **8** to obtain antagonists **9**, **10**, and **11**, respectively.

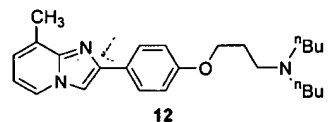


\*Corresponding author. Fax: +1-858-450-2049; e-mail: ncarruth@prdus.jnj.com

Recognizing the deficiencies of the existing imidazole based ligands and the need to identify additional non-imidazole small molecule lead compounds, we undertook a high throughput screening (HTS) approach using the recombinant human receptor. Thus, HTS was initiated with the objective of finding small molecule leads that could ultimately afford compounds that were orally bioavailable and had good BBB penetration. A secondary objective was to evaluate a H<sub>3</sub> antagonist in a range of central nervous system (CNS) disorders.

### Results

To our satisfaction, HTS afforded several lead series with good affinity for the human H<sub>3</sub> receptor including a series of 2-phenyl-imidazo[1,2-*a*]pyridines. These compounds were first prepared as calcium channel blockers and subsequently found to exhibit local anesthetic properties independent of their calcium blocking activity (e.g., RWJ-20085, **12**).<sup>21,22</sup> Thus, **12** was found to be a weak histamine H<sub>3</sub> receptor ligand ( $K_i = 4 \mu\text{M}$ ).

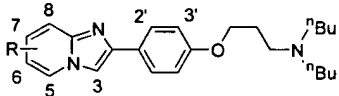


Screening analogues of **12** retaining the di-*n*-butylamino terminus, which had been essential for local anesthetic activity, indicated that a range of substituents were tolerated in the 2-phenyl-imidazo[1,2-*a*]pyridine fragment with minimal effect on potency (Table 1).

However, when we turned our attention to analogues in which the size of the terminal amino group was reduced a dramatic increase in potency was observed. In particular, analogues containing cyclic amines provided significantly more potent compounds (Table 2).

Thus these early screening results quickly demonstrated the favorable effect of a piperidinylpropyl side chain and

**Table 1.** Imidazopyridine and aryl ring substituents, di-*n*-butylamino terminus

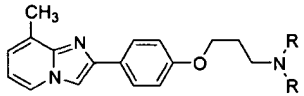
					
No.	R	$K_i$ (nM)	No.	R	$K_i$ (nM)
<b>12</b>	8-CH <sub>3</sub>	2700	<b>20</b>	6,8-di-Br	4800
<b>13</b>	H	1300	<b>21</b>	8-PhCH <sub>2</sub> O	2400
<b>14</b>	7-CH <sub>3</sub>	1100	<b>22</b>	6-Br	2900
<b>15</b>	6-CH <sub>3</sub>	1900	<b>23</b>	8-OH	1500
<b>16</b>	5-CH <sub>3</sub>	2100	<b>24</b>	2',8-di-CH <sub>3</sub>	4200
<b>17</b>	3,8-di-CH <sub>3</sub>	870	<b>25</b>	3',8-di-CH <sub>3</sub>	8000
<b>18</b>	5,7-di-CH <sub>3</sub>	2000	<b>26</b>	3'-OCH <sub>3</sub> , 8-CH <sub>3</sub>	1700
<b>19</b>	8-NO <sub>2</sub>	11,000	<b>27</b>	3',5'-di-OCH <sub>3</sub> , 8-CH <sub>3</sub>	3300

$K_i$  values are the mean of two to six determinations and were determined in house and calculated according to Cheng and Prusoff<sup>23</sup> where  $K_i = 1C_{50}/(1 + [S]/K_d)$  where  $[S] = 0.8 \text{ nM}$  and  $K_d = 0.8 \text{ nM}$  for [<sup>3</sup>H]-*N*-methylhistamine.

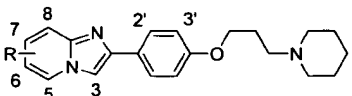
so a limited number of substitutions in the 2-phenyl-imidazopyridine nucleus were examined whilst maintaining the piperidinylpropyl side chain. This indicated that small alkyl groups were tolerated in the imidazopyridine nucleus (Table 3). Simultaneously we observed that the cyclic amine could be piperidine, pyrrolidine or cycloheptylamine (Table 4).

At this juncture, we selected the 7-methylimidazopyridine analogue **38** for more detailed evaluation. The 7-methyl substituent was retained since it was known to reduce both calcium channel affinity and local anesthetic activity in the original series of compounds.<sup>21</sup> Syntheses of the compounds were accomplished

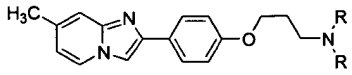
**Table 2.** Amino group substitutions

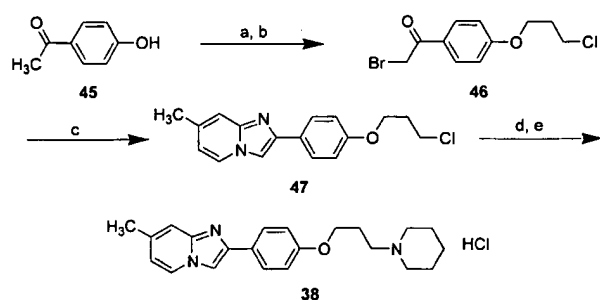
					
No.	NR <sub>2</sub>	$K_i$ (nM)	No.	NR <sub>2</sub>	$K_i$ (nM)
<b>28</b>	N-( <sup>n</sup> C <sub>5</sub> H <sub>11</sub> ) <sub>2</sub>	7900	<b>34</b>	-N<img alt="imidazole ring" data-bbox="785 380 835 405"/>	1000
<b>12</b>	N-( <sup>n</sup> C <sub>4</sub> H <sub>9</sub> ) <sub>2</sub>	2700	<b>35</b>	-N<img alt="morpholine ring" data-bbox="785 415 835 440"/>	80
<b>30</b>	N-( <sup>n</sup> C <sub>3</sub> H <sub>7</sub> ) <sub>2</sub>	43			
<b>31</b>	N-(CH <sub>3</sub> ) <sub>2</sub>	13			
<b>32</b>	N-(CH(CH <sub>3</sub> )Ph) <sub>2</sub>	4000	<b>36</b>	-N<img alt="piperidine ring" data-bbox="785 450 835 475"/>	3
<b>33</b>	NHCH <sub>2</sub> Ph	5400			

**Table 3.** Imidazopyridine and aryl ring substituents, piperidino terminus

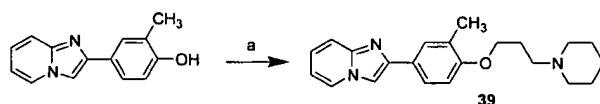
					
No.	R	$K_i$ (nM)	No.	R	$K_i$ (nM)
<b>37</b>	H	6	<b>40</b>	2',7-di-CH <sub>3</sub>	2
<b>36</b>	8-CH <sub>3</sub>	3	<b>41</b>	2'F, 7-CH <sub>3</sub>	10
<b>38</b>	7-CH <sub>3</sub>	2	<b>42</b>	3'-OCH <sub>3</sub> , 7-CH <sub>3</sub>	69
<b>39</b>	3'-CH <sub>3</sub>	28			

**Table 4.** Alkylamine ring size

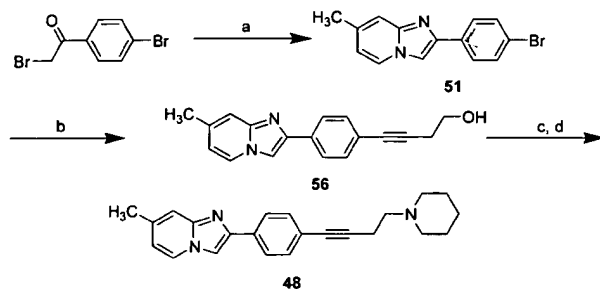
		
No.	-NR <sub>2</sub>	$K_i$ (nM)
<b>38</b>	-N<img alt="piperidine ring" data-bbox="675 815 725 840"/>	2
<b>43</b>	-N<img alt="pyrrolidine ring" data-bbox="675 850 725 875"/>	5
<b>44</b>	-N<img alt="cycloheptylamine ring" data-bbox="675 885 725 910"/>	6



**Scheme 1.** Reagents and conditions: (a)  $K_2CO_3$ , 1-bromo-3-chloropropane, acetone, reflux, 18 h, 98%; (b)  $Br_2$ ,  $Et_2O$ , 18 h, 98%; (c) 2-amino-4-picoline,  $EtOH$ , 73 °C, 2 h, 63%; (d) piperidine, 100 °C, 1.5 h, 90%; (e)  $MeOH$ , 2M  $HCl/Et_2O$ , 100%.



**Scheme 2.** Reagents and conditions: (a) 3-piperidinyl propan-1-ol, DEAD, polymer bound  $PPh_3$ , THF, 20 h, 25%.

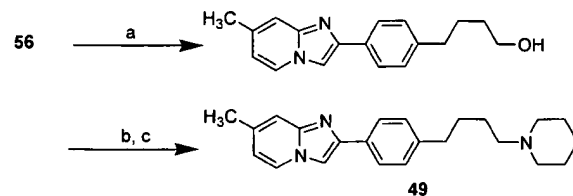


**Scheme 3.** Reagents and conditions: (a) 2-amino-4-picoline,  $EtOH$ , reflux, 18 h, 58%; (b) 3-butyne-1-ol,  $CuI$ ,  $NEt_3$ ,  $Pd(PPh_3)_4$ ,  $CH_3CN$ , reflux, 18 h, 56%; (c)  $MsCl$ , pyridine,  $CH_2Cl_2$ , 80%; (d) piperidine,  $K_2CO_3$ ,  $CH_3CN$ , reflux, 18 h, 97%.

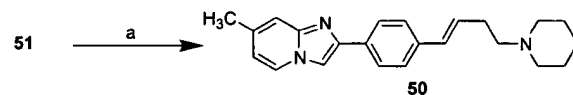
according to the procedure outlined in Scheme 1 for the preparation of **38**. Thus an appropriately substituted 4-hydroxyacetophenone (**45**) was alkylated with 1-bromo-3-chloropropane and brominated to give **46**. Condensation of **46** with a 2-aminopicoline gave **47**, which was treated with an amine to afford **38**.

The aminoalkyl side chain could also be introduced via a Mitsunobu reaction as shown for the preparation of **39** (Scheme 2).

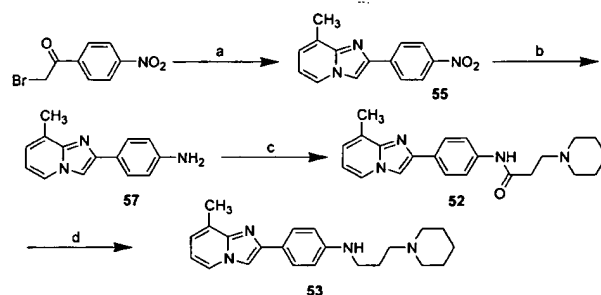
In parallel to the detailed biological evaluation of **38** alternatives to the ether linkage present in **38** were explored (Table 5). The syntheses of these analogues were accomplished as shown in Schemes 3–7. Thus the carbon linked analogues **48–50** were prepared from the 2-(4-bromophenyl)-imidazo[1,2-a]pyridine **51** via either a Sonogashira coupling (Schemes 3 and 4) or a Heck coupling reaction (Scheme 5). The nitrogen linked analogues, **52–54** were prepared from the 2-(4-nitrophenyl)imidazo[1,2-a]pyridine **55** (Scheme 6). The intermediates **51** and **55** were in turn prepared via condensation of an appropriately substituted acetophenone with a 2-amino-picoline.



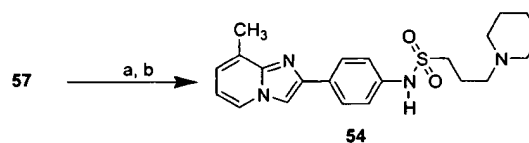
**Scheme 4.** Reagents and conditions: (a) 5%  $Pd-BaSO_4$ ,  $H_2$ ,  $EtOH$ , 4.5 h, 100%; (b)  $MsCl$ ,  $NEt_3$ ,  $CH_2Cl_2$ , 18 h, 100%; (d) piperidine,  $K_2CO_3$ ,  $CH_3CN$ , reflux, 18 h, 48%.



**Scheme 5.** Reagents and conditions: (a) 1-but-3-enylpiperidine,  $Pd(OAc)_2$ ,  $PPh_3$ ,  $NEt_3$ ,  $DMF$ , 150 °C, 20 h, 14%.



**Scheme 6.** Reagents and conditions: (a) 2-amino-3-picoline,  $EtOH$ , reflux, 1.5 h, 22%; (b) cyclohexadiene,  $Pd/C$ ,  $EtOH$ , reflux, 2 h, 50%; (c) 1-piperidinepropionic acid, EDAC, HOBT,  $^iPr_2NEt$ ,  $DMF$ , 18 h, 24%; (d) 2M  $BH_3/DMS$ ,  $PhCH_3$ , reflux, 18 h, 51%.



**Scheme 7.** Reagents and conditions: (a) 3-chloropropanesulfonyl chloride,  $NEt_3$ ,  $DMF$ , 1.5 h, 66%; (b) piperidine, 100 °C, 18 h, 88%.

**Table 5.** Linker variations

No. R = 7-CH <sub>3</sub>	X-Y	K <sub>i</sub> (nM)	No. R = 8-CH <sub>3</sub>	X-Y	K <sub>i</sub> (nM)
<b>38</b>	OCH <sub>2</sub>	2	<b>37</b>	OCH <sub>2</sub>	6
<b>48</b>	C≡C	7	<b>52</b>	NH(CO)	300
<b>49</b>	CH <sub>2</sub> CH <sub>2</sub>	17	<b>53</b>	NHCH <sub>2</sub>	15
<b>50</b>	CH=CH	22	<b>54</b>	NHSO <sub>2</sub> CH <sub>2</sub>	500
<i>trans/cis</i> 9:1					

Although these studies were not exhaustive, they demonstrated that the ether linkage could be replaced with a carbon or an amine linkage without significant loss of affinity. However the amide and sulfonamide analogues **52** and **54** were considerably less active.

## Biological Results and Discussion

Following the successful identification of small molecule histamine  $H_3$  ligands a more detailed biological evaluation was undertaken. Thus in vitro **38** demonstrated high affinity for the human  $H_3$  receptor ( $K_i = 2$  nM) with slightly reduced affinity for the recombinant rat  $H_3$  receptor ( $K_i = 10$ – $20$  nM). In addition **38** exhibited approximately 1000-fold selectivity over the  $H_1$ ,  $H_2$ , and  $H_4$  receptors. Broad screening against a panel of over 50 receptor targets representing the major classes of biogenic amine and neuropeptide receptors, ion channels and neurotransmitter transporters indicated affinities greater than  $K_i = 1$   $\mu$ M. Functional activity versus the human  $H_3$  receptor was determined using SKNMC cells stably transfected with the human  $H_3$  receptor. These compounds, exemplified by **38**, produced a rightward shift in the histamine dose–response curve yielding a  $pA_2 = 8.69$ . The corresponding rat  $pA_2$  was weaker at 8.33. In vivo blood–brain barrier penetration (BBB) was measured following peripheral administration. Thus, **38** was administered (10 mg/kg ip) to rats which afforded a brain  $C_{max} = 10.8 \pm 0.8$   $\mu$ M on a weight/volume basis. Brain receptor occupancy was determined via ex-vivo autoradiography and **38** exhibited an  $ED_{50} = 0.2$  mg/kg sc using [ $^3H$ ]- $R$ - $\alpha$ -methylhistamine to define sites not blocked by the test compound. In the same protocol thioperamide (**3**) had an  $ED_{50} = 2$  mg/kg. Single dose rat pharmacokinetics confirmed that **38** had good oral bioavailability (57%) with a moderate half life ( $t_{1/2} = 5.2 \pm 1.2$  h). Thus, with the objective of identifying small molecule  $H_3$  antagonists having good oral bioavailability and favorable BBB penetration achieved, pharmacological evaluation was initiated in a range of behavioral and cognitive models, details of which will be reported elsewhere.

In summary, proceeding from the cloning of the human  $H_3$  receptor and initiation of a HTS screen a series of novel histamine  $H_3$  ligands were identified. The SARs for the leads were promptly delineated to obtain potent, orally bioavailable, brain penetrating compounds that will aid in the elucidation of the role of central histamine  $H_3$  receptors and determine the therapeutic potential of  $H_3$  receptor antagonists.

## Acknowledgements

The authors are grateful to Ms. Paku Desai for the determination of  $H_4$  receptor affinity and to Dr. Xavier Langlois, Johnson and Johnson Pharmaceutical Research and Development, Beerse, Belgium, for performing the brain receptor occupancy studies described in this manuscript.

## References and Notes

- Dale, H. H.; Laidlaw, P. P. *J. Physiol.* **1910**, *41*, 318.
- Dale, H. H.; Laidlaw, P. P. *J. Physiol.* **1911**, *43*, 182.
- Black, J. W.; Duncan, W. A. M.; Durant, G. J.; Ganellin, C. R.; Parsons, M. E. *Nature* **1972**, *236*, 385.
- Arrang, J.-M.; Garbarg, M.; Schwartz, J.-C. *Nature* **1983**, *302*, 832.
- Hill, S. J.; Ganellin, C. R.; Timmerman, H.; Schwartz, J.-C.; Shankley, N. P.; Young, J. M.; Schunack, W.; Levi, R.; Haas, H. L. *Pharmacol. Rev.* **1997**, *49*, 253.
- Oda, T.; Morikawa, N.; Saito, Y.; Masuho, Y.; Matsumoto, S.-I. *J. Biol. Chem.* **2000**, *275*, 36781.
- Liu, C.; Ma, X.-J.; Jiang, X.; Wilson, S. J.; Hofstra, C. L.; Blevitt, J.; Pyati, J.; Li, X.; Chai, W.; Carruthers, N.; Lovenberg, T. W. *Mol. Pharmacol.* **2001**, *59*, 420.
- Nguyen, T.; Shapiro, D. A.; George, S. R.; Setola, V.; Lee, D. K.; Cheng, R.; Rauser, L.; Lee, S. P.; Lynch, K. R.; Roth, B. L.; O'Dowd, B. F. *Mol. Pharmacol.* **2001**, *59*, 427.
- Zhu, Y.; Michalovich, D.; Wu, H.-L.; Tan, K. B.; Dytko, G. M.; Mannan, I. J.; Boyce, R.; Alston, J.; Tierney, L. A.; Li, X.; Herrity, N. C.; Vawter, L.; Sarau, H. M.; Ames, R. S.; Davenport, C. M.; Hieble, J. P.; Wilson, S.; Bergsma, D. J.; Fitzgerald, L. R. *Mol. Pharmacol.* **2001**, *59*, 434.
- Morse, K. L.; Behan, J.; Laz, T. M.; West, R. E., Jr.; Greenfeder, S. A.; Anthes, J. C.; Umland, S.; Wan, Y.; Hipkin, R. W.; Gonsiorek, W.; Shin, N.; Gustafson, E. L.; Qiao, X.; Wang, S.; Hedrick, J. A.; Greene, J.; Bayne, M.; Monsma, F. J., Jr. *J. Pharmacol. Exp. Ther.* **2001**, *296*, 1058.
- Lovenberg, T. W.; Roland, B. L.; Wilson, S. J.; Jiang, X.; Pyati, J.; Huvar, A.; Jackson, M. R.; Erlander, M. G. *Mol. Pharmacol.* **1999**, *55*, 1101.
- The Histamine  $H_3$  Receptor*; Leurs, R.; Timmerman, H., Eds. Elsevier: Amsterdam, 1998.
- Garbarg, M.; Tuong, M. D. T.; Gros, C.; Schwartz, J.-C. *Eur. J. Pharmacol.* **1989**, *154*, 1.
- Alves-Rodrigues, A.; Leurs, R.; Wu, T.-S.; Prell, G. D.; Foged, C.; Timmerman, H. *Br. J. Pharmacol.* **1996**, *118*, 2045.
- Kathman, M.; Schlicker, E.; Gothert, M. *Psychopharmacology* **1994**, *116*, 464.
- Pompni, S. A.; Gullo, V. P.; Horan, A. C.; Patel, M. G.; Coval, S. US Patent 5,352,707, 1994, *Chem. Abstr.* **1994**, 121, 272182.
- Leurs, R.; Vollinga, R. C.; Timmerman, H. *Prog. Drug Res.* **1995**, *45*, 107.
- Walczynski, K.; Guryn, R.; Zuiderweld, O. P.; Timmerman, H. *Il Farmaco* **1999**, *54*, 684.
- Linney, I. D.; Buck, I. M.; Harper, E. A.; Kalindjian, B. S.; Pether, M. J.; Shankley, N. P.; Watt, G. F.; Wright, P. T. *J. Med. Chem.* **2000**, *43*, 2362.
- Ganellin, C. R.; Leurquin, F.; Piripitsi, A.; Arrang, J.-M.; Garbarg, M.; Ligneau, X.; Schunack, W.; Schwartz, J.-C. *Arch. Pharm. Med. Chem.* **1998**, *331*, 395.
- Sanfilippo, P. J.; Urbanski, M.; Press, J. B.; Dubinsky, B.; Moore, J. B., Jr. *J. Med. Chem.* **1988**, *31*, 2221.
- Dubinsky, B.; Shriver, D. A.; Sanfilippo, P. J.; Press, J. B.; Tobia, A. J.; Rosanthale, M. E. *Drug Dev. Res.* **1990**, *21*, 277.
- Cheng, Y.-C.; Prusoff, W. H. *Biochem. Pharmacol.* **1973**, *22*, 3099.